

Neurophysiological Regulation of Cardiovascular Parameters by the Renin-Angiotensin System: A Review

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ABSTRACT

Cardiovascular diseases and complications form the primary source of deaths globally. The renin-angiotensin system is a ubiquitous system of neuropeptides best known for controlling cardiovascular parameters. This review aims to collect all recent updates on this system in control of heart rate and blood pressure, more specifically, it will tackle the classical and the non-classical renin-angiotensin system and their clinical relevance. To do this, a precise search strategy on the topic will be held on certain databases to find reliable articles and collect the updates. These cardiovascular parameters were found to be controlled by autonomic and neuroendocrine pathways, both of which are initiated from the paraventricular nucleus. The paraventricular nucleus is the site of integration of all peripheral signals from angiotensin II and its further catalytic products. These signals are perceived by the circumventricular organs and the hypothalamic arcuate nucleus through certain receptors and signaling cascades. This system has shown a presence in several diseases and represents a remarkable therapeutic target.

Keywords: Renin-Angiotensin System; Heart Rate; Blood Pressure; Circumventricular Organs; Paraventricular Nucleus; Cardiovascular Neurophysiology

Introduction

The cardiovascular system is a highly regulated blood-pumping heart and vessels. Dysregulation in its functioning is frequent and would lead to many cardiovascular diseases (CVDs). According to the world health organization (WHO), CVDs are the leading cause of death globally, forming up to 32% of all global deaths [1]. Among CVDs, unachieved homeostasis of cardiovascular parameters is an uprising issue to be addressed. Studies have linked the control of cardiovascular parameters to the autonomic nervous system (ANS) and several neuropeptides, among which the renin-angiotensin-aldosterone system (RAAS or RAS) provides many neuroactive forms of angiotensin peptides involved in controlling the ANS and peripheral neurotransmission [2]. RAS has been recognized for its importance in regulating blood pressure homeostasis, extracellular fluid volume, and the neural and endocrine functions in cardiovascular control, hence its role in many pathophysiological CVDs including hypertension, heart failure, coronary artery disease, strokes, and cardiovascular-related diseases as obesity and chronic kidney disease [3].

The RAS is a ubiquitous hormonal system that includes a series of enzymatic reactions, peptide hormones, and byproducts. The classic RAS is a systemically acting system in the bloodstream. Other renin-angiotensin systems are tissue-specific in the brain, kidney, heart, blood vessels, adipocytes, and adrenal glands; they operate completely independent of the

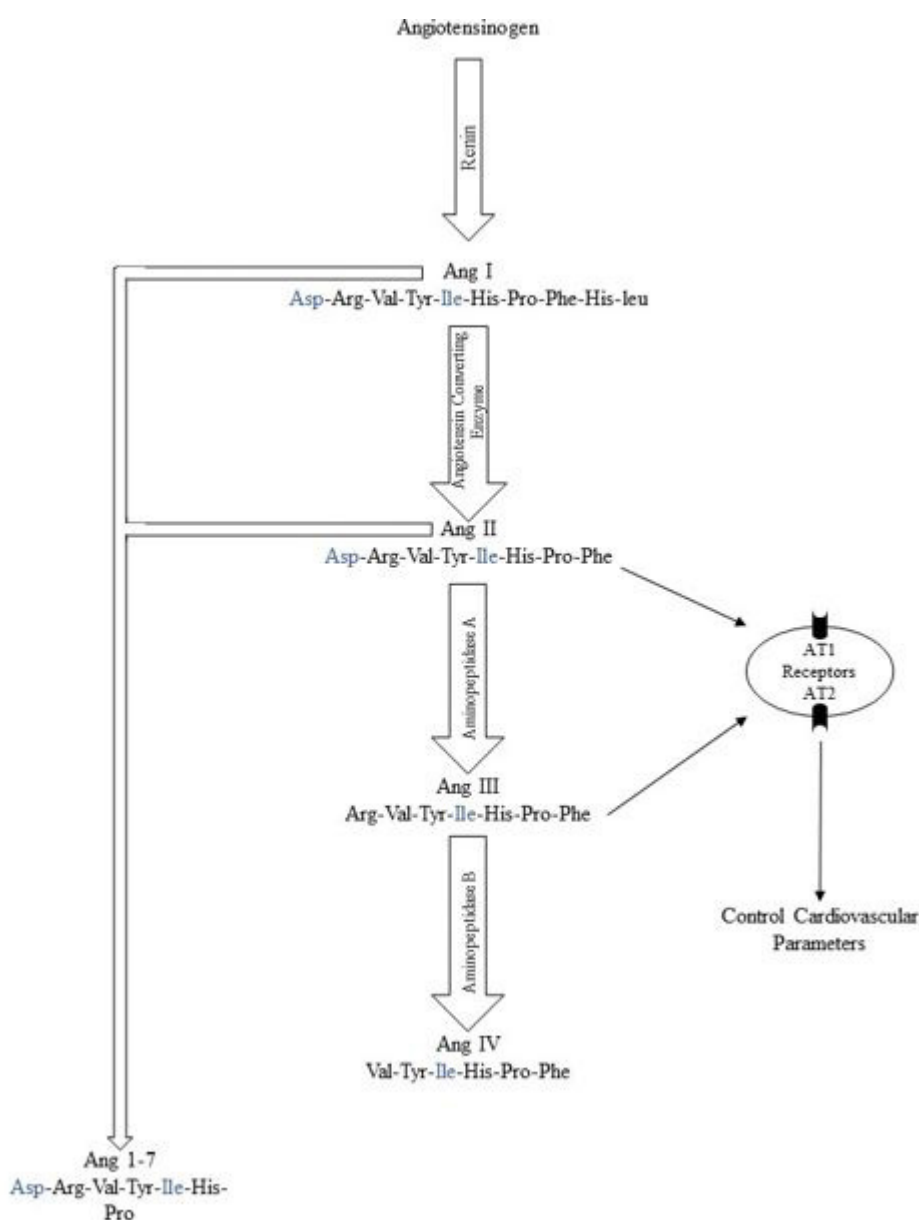


Figure 1: Illustration on the enzymatic and peptide series of the renin-angiotensin system. Angiotensinogen is catalyzed by renin into Ang I, which is then catalyzed into Ang II by angiotensin converting enzyme. Ang II can be further catalyzed into Ang III and Ang IV by aminopeptidase A and B, respectively. Ang 1-7 is a product of catalysis of Ang I and Ang II. Ang II and Ang III have the same functions and act on the same receptors AT1 and AT2. Ang: angiotensin, AT1/2: angiotensin II receptor type 1/2. (Arab, 2013)

classic RAS except for renin and angiotensinogen, which can only be obtained from the bloodstream [4]. Other RAS can also be intracellular as in cardiac myocytes, vascular smooth muscle cells, renal proximal tubule cells, and neurons; in these systems, angiotensin II, the representative neuropeptide of RAS, can either be synthesized or internalized [5]. Generally, this series of enzymatic starts with angiotensinogen cleavage into angiotensin I by renin, then into angiotensin II (Ang II) by angiotensin-converting enzyme. Furthermore, Ang II is cleaved into angiotensin III (Ang III) by aminopeptidase A and then angiotensin IV (Ang IV) by aminopeptidase N. Another cleavage series of Ang II involves the production of angiotensin 1-7 (Ang 1-7), which can be further cleaved into alamandine [6-7]. Ang II exerts its activity through interactions with its receptors angiotensin II receptor types 1 and 2 (AT1R/AT2R), Ang III has the same effects and same receptor compatibility to Ang II (Figure 1) but it is five times faster cleared from the circulation [8]. Ang IV, however, interacts with angiotensin II receptor type 4 (AT4R) but has rather far roles from the regulation of blood pressure and heart rate [9].

The research on RAS and its link to heart rate and blood pressure control are intensive and only a few reviews have considered covering this topic, or at least parts of it. This review will tackle most of the details related to this topic and provide an in-depth collection of recent advances in this field. Hence, RAS, here, will be classified into:

Classical RAS: this involves a brief introduction to Ang II production and its activity on the lamina terminalis in the brain, and how recent studies have tackled them.

Another part of this classification in the non-classical RAS: it involves the novel Ang 1-7, alamandine, and the prorenin receptor – a receptor for prorenin and/or renin that is known to non-proteolytically cleave angiotensinogen to produce angiotensin I -, in addition to proinflammatory molecules as tumor necrosis factor α and the interleukins.

Finally, RAS is to be applied in the clinical field of research.

Classical Renin-Angiotensin System and the Brain

The classic RAS acts systemically and begins in the juxtaglomerular cells of the renal afferent arteriole of the kidney. These cells synthesize prorenin that is cleaved to renin in the endoplasmic reticulum and packaged into secretory granules in the Golgi apparatus [10]. While prorenin is constitutively secreted across the cell membrane, mature renin is only released upon a stimulus by the afferent arteriolar baroreceptor when arterial perfusion pressure decreases or when the sympathetic nervous system is activated and stimulates norepinephrine release and activity on beta-1 adrenergic receptors [11]. Renin is responsible for the cleavage of angiotensinogen, a precursor majorly produced by the liver and secreted into the bloodstream, into angiotensin I (Ang I). Ang I fate is to be converted by the angiotensin-converting enzyme (ACE) into angiotensin II (Ang II), the representative active neuropeptide of RAS [12].

Central angiotensin II (Ang II) has been known to elevate blood pressure as early as 1963 [13]. It interacts with angiotensin II type 1 receptor (AT1R) on the circumventricular organs (CVOs): the subfornical organ (SFO), the median preoptic nucleus (MnPO), and the organ vasculosum of the lamina terminalis (OVLT). These CVOs are blood-brain barrier deficient and act as the interface between blood and its signals to the brain, hence the role as sensory organs. They receive the signal and transduce it to excitatory - through glutamate neurotransmitter - or inhibitory - through γ -aminobutyric acid (GABA) - projections towards the hypothalamic paraventricular nucleus (PVN) either directly or indirectly through the MnPO [14-15-16].

Subfornical Organ (SFO)

A 40-year-old notable study on the SFO has reported it as a source of producing Ang II in addition to a sensory organ [16-17-18]. Later in 2017, this was addressed again. The 2017 study has characterized a superficial population of secretory cells on the SFO that are similar to other SFO neurons except for the secretory vesicles within. Real-life monitoring of Vesicle Associated Membrane Protein 2 (VAMP2) confirmed their exocytosis and uncovered their route into the ventricles. This SFO secretory role may be providing a direct route for the effects mediated by SFO on blood pressure and heart rate [19].

Paraventricular Nucleus (PVN)

The PVN is of major interest to the researchers in this field for it acts as an integrative center, receiving projections from different brain regions including the CVOs and arcuate nucleus, and sending projections to the hypothalamus-pituitary-adrenal (HPA) axis and pre-sympathetic projections to the rostral ventrolateral medulla (RVLM) [20].

The paraventricular nucleus is functionally divided into two parts: the magnocellular PVN (PVNm) and the parvocellular PVN (PVNp). Afferent Glutamatergic projections increasing the firing of neurons within the PVN is the basic knowledge, however, several recent findings were reported in the last 5 years [21].

Parvocellular Paraventricular Nucleus

PVNp neurons are presympathetic projections towards the RVLM responsible for adjusting the autonomic nervous messages towards the heart, vessels, and kidneys to achieve homeostasis [22].

Within the PVNp, a mutual activating system between several angiotensinergic (Ang) neurons and vasopressinergic (VP) neurons functions to deliver these adjustments [23]. The latter responds to excitatory signals from the former through AT1R and angiotensin II type 2 receptor (AT2R), though AT2R accounts for the main receptor [24]. Now that VP neurons are activated, they secrete vasopressin to Ang neurons, which, in turn, respond through their V1a receptor. However, the mutual activating system might cause an over-activation, this requires GABAergic interneurons activation; Ang neurons send excitatory projections to GABAergic interneurons soma and presynaptic terminals acting on AT2R and AT1R, respectively, and these neurons send inhibitory projections to the VP neurons. Moreover, activated VP neurons send the pre-sympathetic projections to the RVLM [25] (Figure 2).

Magnocellular Paraventricular Nucleus

The PVNm are involved in the neuroendocrine pathway, extending their projections to the pituitary gland. Angiotensin II signals received by the CVOs activate the vasopressinergic neurons within, which then act to activate the pituitary gland [26] (Figure 2).

A Cross-Talk between the Autonomic and Neuroendocrine Control of Cardiovascular Parameters

Within the PVN, some neurons have been reported to express AT1aR. Selective optogenetic activation or inhibition of these neurons, respectively, activate or inhibit the HPA axis; these neurons control the neuroendocrine pathway of cardiovascular parameter control [27]. In a subsequent study, it was detected that these neurons express corticosteroid-releasing hormone (CRH) and make appositions with the presympathetic neurons sent from the PVN to the RVLM and control the sympathetic activity. The former neurons respond to CRH secretion through the CRH type 1 receptor (CRHR1) and transduce the signal as an excitatory message [28] (Figure 2)

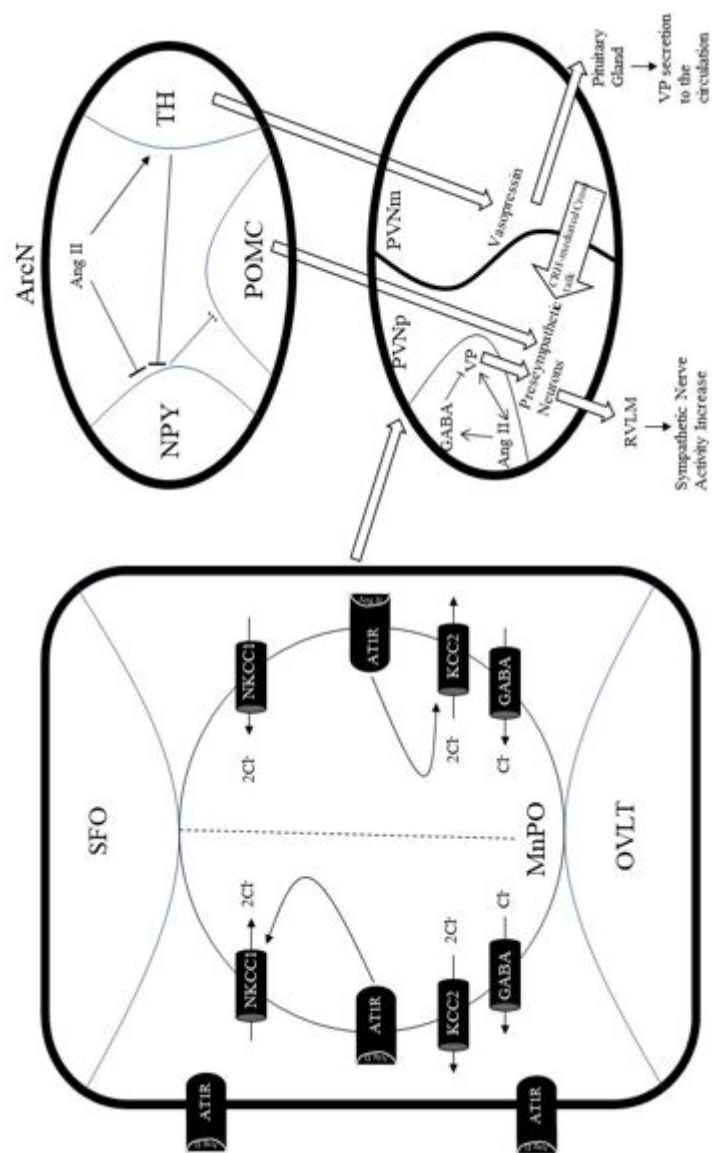


Figure 2: An illustration on the molecular mechanisms that occur within the brain regions involved in RAS signaling. Within the MnPO, depending on AT1R signal, it favors NKCC1 in normal conditions or KCC2 in dysregulations, which only causes the projections towards the PVN to be inhibitory or lessinhibitory, respectively. Within ArcN, Ang II causes a disinhibition of the POMC either through direct actions on NPY neurons or indirectly through TH neurons; hence, POMC activates presympathetic neurons of the PVNp and TH neurons activate vasopressin neurons in the PVNm. Within the PVN, a mutual activating system between angiotensinergic and vasopressinergic neurons in the PVNp activates the presympathetic neurons, activated neurons in the PVNm activate the pituitary through vasopressin and the presympathetic neurons through CRH as means of cross-talking. Ang II: angiotensin II, AT1R: angiotensin II receptor type 1, NKCC1: Na-K-Cl cotransporter isoform 1, KCC2: -Cl cotransporter, isoform 2, GABA: γ -aminobutyric acid, SFO: subfornical organ, MnPO: median preoptic nucleus, OVLT: organ vasculosum of the lamina terminalis, ArcN: arcuate nucleus, POMC: proopiomelanocortin, TH: tyrosine hydroxylase, NPY: neuropeptide Y, PVNp: parvocellular paraventricular nucleus, PVNm: magnocellular paraventricular nucleus, VP: vasopressin, CRH: corticotropin-releasing hormone, RVLM: rostral ventrolateral medulla. (Elsaafien et al., 2021; Farmer, Little, Marcianite, & Cunningham, 2021; Frazier et al., 2021; Nasimi, Haddad, Mirzaei-Damabi, Rostami, & Hatam, 2021; Shi, Stornetta, Stornetta, & Brooks, 2022)

Arcuate Nucleus and RAS

Studies on Ang II and the arcuate nucleus (ArcN) first come up in 2007, this study showed increased activity of the ArcN in response to Ang II [29]. Later in 2011, the ArcN has shown to elevate the arterial blood pressure in response to Ang II, and hence, its increased activity [30]. Only recently, the mechanism of action of Ang II on the ArcN has been deciphered. The arcuate nucleus has several populations of neurons, some of our interests here are the proopiomelanocortin (POMC) -expressing neurons, tyrosine hydroxylase (TH) - expressing interneurons, and neuropeptide Y (NPY) - expressing neurons [31].

In the absence of the Ang II signal, NPY neurons send inhibiting projections to POMC and PVN neurons acting through Y1 receptors. Ang II inhibits NPY neurons either directly through inhibitory AT1aR on NPY neurons or indirectly through activating AT1aR receptors on TH neurons that send GABAergic projections to NPY neurons. This causes disinhibition on the POMC neurons in particular, which, then, activates PVN neurons expressing melanocortin-4-receptor (MC4R) through secreting α -Melanocyte-stimulating hormone (α -MSH). TH neurons might also activate the pituitary gland either directly or indirectly through activating projections to vasopressinergic neurons in the PVN. This works as a 2-phase response to the Ang II signal in the arcuate nucleus: it first starts with increased vasopressin secretion to the circulation and ends with increased sympathetic activity from the PVN [32] (Figure 2).

Median Preoptic Nucleus

Researchers have been studying MnPO as a site of dysregulation in RAS signaling. Early studies have only linked MnPO to ACE to dysregulations of RAS signaling [33-34-35]. Putting pieces together, subsequent studies started linking it to Ang II and glutamate signaling [36]. And, only recently, a decisive mechanism was established. This dysregulation is due to an imbalance between excitatory and inhibitory messages; the former increase and the latter decreases. The increased activation of AT1aR causes a functional shift in the activity of chloride channels - Na-K-2Cl cotransporter isoform 1 (NKCC1) influx ion channels and K-Cl cotransporter isoform 2 (KCC2) efflux ion channels. Initially, normal AT1aR activity maintains a low level of chloride ions within the neurons as it favors the activity of KCC2, hence GABA receptors are activated and transport chloride ions into the neurons leading to the formation of an inhibitory message. However, increased activity of AT1aR favors NKCC1 leading to high levels of intracellular chloride ions, and, eventually, GABA receptors act to expel chloride ions leading to less inhibition or even excitation in some cases [37] (Figure 2).

Central RAS at the Molecular Level

The interaction of Ang II with its receptor AT1R leads to a glutamatergic message. Glutamate interacts with either alpha-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA) GluA1 receptor [38] or N-methyl-D-aspartate (NMDA) receptor [38]. This interaction either causes an influx of calcium ions into the neuron [39] or activates protein kinase C δ (PKC δ) [40], which, in both cases, interacts with nicotinamide adenine dinucleotide phosphate (NADPH) oxidases [41], specifically through Ras-related C3 botulinum toxin substrate 1 (Rac1) in case of PKC δ [42]. This only leads to the production of reactive oxygen species (ROS), especially superoxides [43]. Potassium channels might be regulated by superoxides mediated by calcium/calmodulin (CaM)-dependent kinase II α (CaMKII α) as a redox-sensitive signaling protein [44] (Figure 3).

In another study, Ang II response was reported to be related to brain cytosolic phospholipase alpha 2 (cPLA2). Activated cPLA2 leads to the production of arachidonic acid that is catalyzed by cyclooxygenase 2 into prostaglandin E2 (PGE2). PGE2 binds its receptors Prostaglandin E2 receptor 2/4 (EP2/4) to mediate RAS effects [45] (Figure 3).

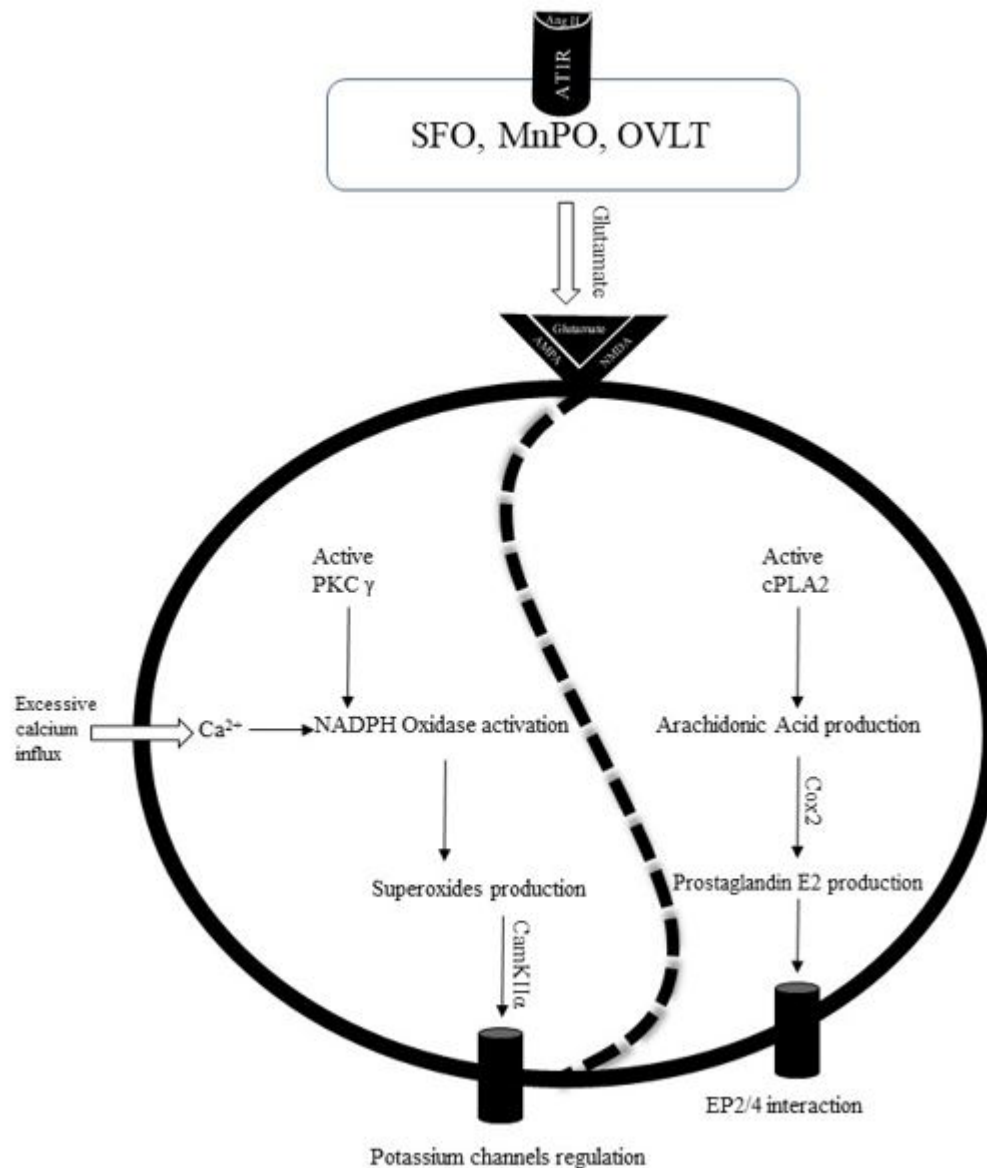


Figure 3: An illustration on the two molecular pathways of Ang II signaling. Ang II-AT1R interaction in SFO, OVLT, and MnPO induces glutamate signaling that acts on both AMPA and NMDA receptors of glutamate leading to molecular mechanisms within the cells of the paraventricular nucleus. The first scenario shows excessive calcium influx and active PKC gamma that activate NADPH oxidation further leading to superoxide production and regulating potassium channels through CamKII alpha. A second scenario shows active cPLA2 producing arachidonic acid, which is catalyzed into prostaglandin E2 by Cox2, and the latter interacts with EP2/4 for further responses. Ang II: angiotensin II, AT1R: angiotensin II receptor type 1, SFO: subfornical organ, MnPO: median preoptic nucleus, OVLT: organ vasculosum of the lamina terminalis, PKC: protein kinase C, NADPH: reduced nicotinamide adenine dinucleotide phosphate, CamKII alpha: calcium/calmodulin (CaM)-dependent kinase II α , cPLA2: brain cytosolic phospholipase alpha 2, Cox2: cyclooxygenase 2, EP2/4: prostaglandin E2 receptor 2/4. (Basu et al., 2019; Song et al., 2018; Su et al., 2017)

Non-classical Renin-Angiotensin System and the Brain

For many years, Ang II was thought to be the only active neuropeptide in RAS. However, two decades earlier, angiotensin-converting enzyme 2 (ACE2) was discovered [46]. ACE2 is an enzyme that further catalyzes Ang II into angiotensin 1-7 (Ang 1-7), which can be further decarboxylated into alamandine. These relatively new components of RAS along with angiotensin II type 2 receptor (AT2R) fall in the protective arm of RAS considered to be vasodilators and lower the cardiovascular parameters [47]. However, these components have arisen many controversies about their actions.

Angiotensin 1-7 and Alamandine

Most of the functions of Ang 1-7 are mediated through its interaction with mitochondrial assembly protein 1 receptor (MasR) and those of alamandine are mediated through Mas-related receptor member D (MrgD) [48]. The controversy with these RAS components revolves around their position: In the periphery and caudal ventrolateral medulla, they act as a vasodilator – counter-regulating Ang II. However, in RVLM and PVN, they act as vasoconstrictors [49].

Another controversy related to Ang 1-7 is about its molecular pathway. In an earlier study in 2005, Ang 1-7, through its interaction with MasR, was thought to activate the NO pathway [50]. Later in 2017, in a study that showed a depressor effect of Ang 1-7 on the PVN, it was reported that Ang 1-7 exerts its effects through the cyclic adenosine monophosphate (cAMP)-protein kinase A (PKA) pathway [51] and through reduced nicotinamide adenine dinucleotide phosphate (NADPH) oxidase-derived superoxide anions and not through NO pathway [52].

Tumor Necrosis Factor α and RAS

As a pro-inflammatory cytokine, tumor necrosis factor α or TNF α can exist in two forms: a benign and rather beneficial transmembrane TNF α (trTNF α) and a soluble form (sTNF α) [53] known for its inflammatory effects and involved in blood pressure control in some conditions through epidermal growth factor receptor (EGFR) and extracellular signal-regulated kinase (ERK) 1/2 signaling molecules [54]. This signaling cascade starts with TNF α receptor 1 (TNF α R1) [55].

Tumor Necrosis Factor α Receptor 1 and AT1R

TNF α R1 colocalizes with AT1R in SFO neurons [55] and its interaction with its ligand TNF α shifts the activation threshold of voltage-gated sodium channels to a hyperpolarized state rendering the depolarization of these neurons more robust; their excitability increases [56]. This concludes with TNF α potentiating the induced calcium rise in response to Ang II in SFO [57].

ADAM Metallopeptidase Domain 17, AT1R, and ACE2

The molecule responsible for catalyzing the reaction from trTNF α to sTNF α is called TNF α converting enzyme (TACE), also known as ADAM Metallopeptidase Domain 17 (ADAM17). ADAM17 has a rather noteworthy involvement in RAS. It is expressed in the periphery and brain regions as SFO and PVN on neurons and glial cells [58].

While, at first sight, ADAM17 does not show any direct relevance to RAS components, ADAM17 co-exists with AT1R in many cells and can be activated and upregulated by AT1R activation; Once AT1R is activated, both molecules are brought into a closer proximity for downstream signals to activate ADAM17 through intracellular signals from G-protein coupled receptors or oxidative stress that phosphorylate its cytoplasmic domain or upregulate its availability due to translocation in acute stimulations or upregulate its availability through the involvement of transcription factors in chronic stimulations [59].

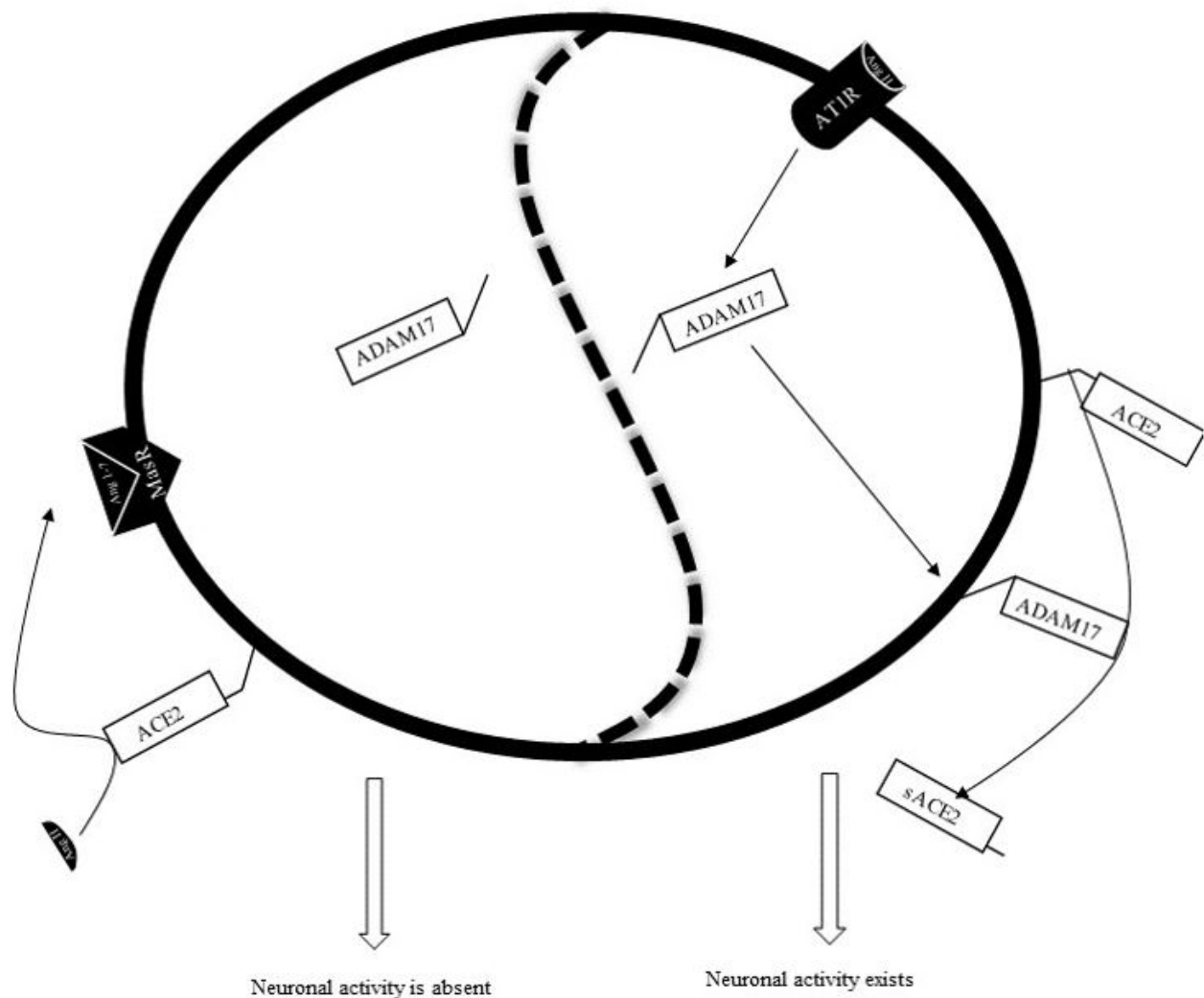


Figure 4: An illustration on how AT1R overactivation modulates ADAM17 activity leading to a neuronal activity and how ACE2 can have a compensatory role in normal conditions. ACE2 converts Ang II into Ang 1-7, which interacts with MasR to produce compensatory effects. In overactive Ang II, AT1R induces colocalization of ADAM17 into the the membrane, which then converts ACE2 into sACE2 leading to an active neuronal response. Ang: angiotensin , ACE2: angiotensin converting enzyme 2, MasR: mitochondrial assembly protein 1 receptor, ADAM17: ADAM Metallopeptidase Domain 17, sACE2: soluble ACE2. (Mukerjee et al., 2019; Xu et al., 2017)

An interesting ligand of ADAM17 is ACE2, in which they interact in a process called ADAM17-mediated shedding of ACE2. This interaction regulates the activity of presympathetic neurons of the PVN: neuronal ACE2 supports an inhibitory input to the PVN, ADAM17 cleaves ACE2 membrane proteins reducing its compensatory effects [60], and assists glutamatergic neurons that induce sympathoexcitation at some point [61]. A controversy, here, lies in the activity of the shed, now soluble, ACE2 in whether it has a therapeutic benefit or it is merely a degradation product [62] (Figure 4).

Interleukins and RAS

Low-grade inflammation is detected in hypertension in the peripheral tissues, blood [63-64], and the cardiovascular centers in the brain [65-66-67-68]. Studies relating proinflammatory cytokines and hypertension has come to mention three main cytokines: Interleukin 1 β , Interleukin 17A, and Interleukin 10.

As for interleukins that induce hypertension, microinjection studies of Interleukin 1 β in the SFO induces hypertension [69]. In a subsequent study, interleukin 1 β in the circulation showed an interaction with the SFO to produce hypertension through inflammation and increasing RAS activity [70]. Moreover, Interleukin 17A is involved in neuroinflammation and sympathetic activation during hypertension. In fact, Interleukin 17A readily crosses the blood-brain barrier to boost inflammation in the brain through various inflammatory mediators in the PVN [71].

Regarding protective interleukins, alterations of the expression of interleukin 10 and its receptor takes place in hypertension [72], suggesting their involvement. In fact, interleukin 10 has a rather protective effect by lowering the low-grade inflammation [73].

Brain Local RAS and the (Pro) renin Receptor

RAS components also exist in tissues. One of the most remarkable components in local RAS is the (pro)renin receptor ((P)RR or PRR). PRR binds to prorenin, the inactive form of renin, and/or renin to catalyze a fourfold increased production of Ang I [74] or a non-proteolytic production of Ang II [75] in response to specific signals as high salt intake [76], directly affect Ang II signaling [77], or increase in the excitability of magnocellular neurons, through increasing intracellular calcium and inhibition A-type potassium channels, leading to an increase in vasopressin secretion in an Ang II-independent pathway [78]. Within the brain, local RAS components have autocrine or paracrine effects [79].

Clinical Relevance of RAS

In context of diseases, RAS has been linked to hypertension as a disease and a complication in many diseases. Hypertension has been linked to overactive Ang II -especially since the blood-brain barrier shows a higher permeability in hypertension [80] in the PVN [81] and ArcN [82], Ang 1-7 [83] and alamandine in the PVN [84], and to the action of ADAM17 corresponding to ACE2 activity [85]. Other brain regions and other related molecules have also been linked to hypertension due to elevated sympathetic activity and excitatory nerve messages in the OVLT and MnPO and the RVLM [86]. Some remarkable molecules are TNF α [87], ADAM17 [88], and PRR [89]. This also falls for hypertension which is a complication of other diseases, specifically talking about chronic intermittent hypoxia (CIH) [90], obesity polycystic kidney disease (Underwood, McMullan, Goodchild, Phillips, & Hildreth, 2022), and stress-related anxiety.

As for treatments that relate to RAS, lesions of MnPO neurons that project to the PVN have been reported viable for treating CIH. Several other less invasive treatments have also been studied; any of the involved molecules in hypertension is likely to be a target: AT1R antagonist losartan has shown to lower blood pressure and heart rate, Ang 1-7 can be selectively blocked by D-Alanine-Ang-1-7 (A-779), in addition to PRR antagonists as PRR knockdown was proven efficient to reduce hypertension. Following the cascade that Ang II receptors activate, the cascade can be a viable therapeutic target antioxidants have, though, proven insufficient for the treatment of hypertension due to reasons related to sublocalization and specificity of ROS type; more specific antioxidants should be directed against superoxides as they have been reported to be elevated during hypertension. Studies on Metformin have also linked it to reducing the oxidative stress related to hypertension.

Conclusion and Future Perspectives

The renin-angiotensin system is a ubiquitous hormonal system with several neuropeptide components acting on numerous targets through various receptors. It has been assigned to several functions; one of our interests is its effect on the cardiovascular parameters: heart rate and blood pressure. This area of research is intensive and most of the reviews on it are rather considering parts of the system as to how massive this system is. In this review, RAS has been briefly described, the recent advances in this field were in-depth collected, and a clinical significance to what RAS can help with was provided.

Studies have shown its involvement in controlling heart rate and blood pressure via the autonomic and neuroendocrine systems, ultimately, increasing the sympathetic nerve activity and activating the pituitary, however, signals are first integrated within the PVN. Signals towards the PVN are initiated within the CVOs, where SFO not only responds to Ang II but also can be its source into the ventricles, and where MnPO utilizes two channels in order to either activate GABA receptor chloride ion channels and, hence, send inhibitory messages to the PVN in case of low Ang II signal or rather be less inhibitory in case of high Ang II signaling. The ArcN also receives Ang II signals and, ultimately, sends excitatory projections to the PVN presympathetic neurons or to vasopressinergic neurons, both of which cross-talk through CRH. The PVN, itself, has a mutual activating system between angiotensinergic and vasopressinergic neurons that ultimately activates the presympathetic neurons. All of this shows how Ang II ultimately increases heart rate and blood pressure through activating the sympathetic nervous and neuroendocrine systems. Ang II metabolites are still controversial

in terms of periphery versus central actions. Moreover, the proinflammatory cytokines and TNF α convertase have shown to be involved in the dysregulations during hypertension.

This collection of RAS updates pinpoints not only how intensive the research is, but also how redundant it is in the case of the PVN especially since it is the site of integration of these peripheral signals. Studies on non-classical RAS have not shown any in-depth results and are still on the margins. In conclusion, RAS is a broad source of neuropeptides acting on many receptors in various regions of the body, and understanding its exhaustive list of activities is a step toward treating many disease-related complications.

With the drugs utilizing RAS in the market: angiotensin-receptor blockers (ARBs or sartans), angiotensin-converting enzyme inhibitors, and direct renin inhibitors (aliskiren), RAS has not been in the clinical field for some time now; in a span of 27 years starting from 1986, only 3 clinical trials were reported related to either brain regions involved of RAS signaling or RAS component Ang II. While current studies still aim to understand the renin-angiotensin system furthermore, future studies should utilize this knowledge and push RAS metabolites and neuropeptides into the clinical field.

Conflict of Interest

The authors declare that they have no conflict of interest.

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References

1. Agassandian K, Grobe J L, Liu X, Agassandian M, Thompsoz A P, Sigmund C D, Cassell M D, et al. (2017) Evidence for intraventricular secretion of angiotensinogen and angiotensin by the subfornical organ using transgenic mice *American journal of physiology Regulatory, integrative and comparative physiology* 312: 973-981.
2. Arab AA (2013) Effets des neuropeptides pancréatiques, des angiotensines et de l'activité physique sur les paramètres cardiovasculaires et respiratoires (PhD), Université de Bretagne Occidentale, *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*.
3. Arakawa H, Chitravanshi VC, Sapru HN (2011) The hypothalamic arcuate nucleus: a new site of cardiovascular action of angiotensin-(1-12) and angiotensin II *Am J Physiol Heart Circ Physiol*, 300: 951-960.
4. Basu U, Case A J, Liu J, Tian J, Li YL, Zimmerman MC, et al. (2019) Redox-sensitive calcium/calmodulin-dependent protein kinase II α in angiotensin II intra-neuronal signaling and hypertension *Redox Biol*, 27: 101230.
5. Brasil TFS, Belém-Filho IJA, Fortaleza EAT, Antunes-Rodrigues J, Corrêa FMA, et al. (2022) The AT-1 Angiotensin Receptor is Involved in the Autonomic and Neuroendocrine Responses to Acute Restraint Stress in Male Rats *Cell Mol Neurobiol*, 42:109-124.
6. Cao Y, Yu Y, Xue B, Wang Y, Chen X, Beltz T G, Wei SG, et al. (2021) IL(Interleukin)-17A Acts in the Brain to Drive Neuroinflammation, Sympathetic Activation, and Hypertension *Hypertension* 78:1450-62.
7. Carey RM, Padia SH (2018) Chapter 1 - Physiology and Regulation of the Renin–Angiotensin–Aldosterone System In A K Singh & G H Williams (Eds), *Textbook of Nephro-Endocrinology (Second Edition)* Academic Press.
8. Case AJ, Tian J, Zimmerman MC (2017) Increased mitochondrial superoxide in the brain, but not periphery, sensitizes mice to angiotensin II-mediated hypertension *Redox Biol*, 11:82-90.
9. Chamarthi B, Williams GH, Ricchiuti V, Srikumar N, Hopkins PN, Luther JM, Thomas A, et al. (2011) Inflammation and Hypertension: The Interplay of Interleukin-6, Dietary Sodium, and the Renin–Angiotensin System in Humans *Am J Hypertens*, 24:1143-8.
10. Bautista LE, Vera LM, Arenas IA, Gamarra G (2005) Independent association between inflammatory markers (C-reactive protein, interleukin-6, and TNF- α) and essential hypertension *Journal of Human Hypertension*, 19:149-54
11. Chertow G, Yu A, Taal M, Skorecki K, Marsden P, Luyckx V, et al. (2019) *Brenner and Rector's The Kidney (Vol 1)* Elsevier: Elsevier.
12. Dampney RA, Michelini LC, Li DP, Pan H L, et al. (2018) Regulation of sympathetic vasomotor activity by the hypothalamic paraventricular nucleus in normotensive and hypertensive states *Am J Physiol Heart Circ Physiol*, 315:00-14.
13. Davern PJ, Head GA (2007) Fos-related antigen immunoreactivity after acute and chronic angiotensin II-induced hypertension in the rabbit brain *Hypertension*, 49:1170-7.
14. de Kloet AD, Wang L, Ludin JA, Smith JA, Pioquinto DJ, Hiller H, Krause EG, et al. (2016) Reporter mouse strain provides a novel look at angiotensin type-2 receptor distribution in the central nervous system *Brain Struct Funct*, 221:891-912.
15. de Kloet AD, Wang L, Pitra S, Hiller H, Smith J A, Tan Y, Krause EG, et al. (2017) A Unique “Angiotensin-Sensitive” Neuronal Population Coordinates Neuroendocrine, Cardiovascular, and Behavioral Responses to Stress *J Neurosci*, 37:3478-90.
16. Dickinson CJ, Lawrence JR (1963) A slowly developing pressor response to small concentrations of angiotensin Its bearing on the

pathogenesis of chronic renal hypertension *Lancet*, 1:1354-6.

17. Donoghue M, Hsieh F, Baronas E, Godbout K, Gosselin M, Stagliano N, Acton S, et al. (2000) A novel angiotensin-converting enzyme-related carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1-9 *Circ Res*, 87:1-9.

18. Elsaafien K, Kirchner MK, Mohammed M, Eikenberry SA, West C, Scott KA, Krause EG, et al. (2021) Identification of Novel Cross-Talk between the Neuroendocrine and Autonomic Stress Axes Controlling Blood Pressure *The Journal of Neuroscience*, 41(21), 4641 doi:10.1523/JNEUROSCI.0251-21.2021

19. Farmer GE, Little JT, Marcianite AB, Cunningham JT (2021) AT1a-dependent GABA(A) inhibition in the MnPO following chronic intermittent hypoxia *American journal of physiology Regulatory, integrative and comparative physiology*, 321:469-81.

20. Faulk K, Shell B, Nedungadi TP, Cunningham JT (2017) Role of angiotensin-converting enzyme 1 within the median preoptic nucleus following chronic intermittent hypoxia *American journal of physiology Regulatory, integrative and comparative physiology*, 312:245-52.

21. Faulk K E, Nedungadi T P, Cunningham J T (2017) Angiotensin converting enzyme 1 in the median preoptic nucleus contributes to chronic intermittent hypoxia hypertension *Physiol Rep*, 5:13277.

22. Frazier C J, Harden S W, Alleyne A R, Mohammed M, Sheng W, Smith J A, de Kloet AD, et al. (2021) An Angiotensin-Responsive Connection from the Lamina Terminalis to the Paraventricular Nucleus of the Hypothalamus Evokes Vasopressin Secretion to Increase Blood Pressure in Mice *J Neurosci*, 41:1429-42.

23. Fry WM, Ferguson AV (2021) The subfornical organ and organum vasculosum of the lamina terminalis: Critical roles in cardiovascular regulation and the control of fluid balance *Handb Clin Neurol*, 180:203-15.

24. Gong J, Shen Y, Li P, Zhao K, Chen X, Li Y, Kong X, et al. (2019) Superoxide anions mediate the effects of angiotensin (1-7) analog, alamandine, on blood pressure and sympathetic activity in the paraventricular nucleus *Peptides*, 118:170101.

25. Iovino M, Messana T, De Pergola G, Iovino E, Guastamacchia E, Licchelli B, Triggiani V, et al. (2020) Brain Angiotensinergic Regulation of the Immune System: Implications for Cardiovascular and Neuroendocrine Responses *Endocr Metab Immune Disord Drug Targets*, 20:15-24.

26. Johnson AK, Epstein AN (1975) The cerebral ventricles as the avenue for the dipsogenic action of intracranial angiotensin *Brain Research*, 86:399-418.

27. Katsurada K, Ogozama Y, Imai Y, Patel KP, Kario K, et al. (2021) Renal denervation based on experimental rationale *Hypertension Research*, 44:1385-94.

28. Khanmoradi M, Nasimi A (2017) Functions of AT1 and AT2 angiotensin receptors in the paraventricular nucleus of the rat, correlating single-unit and cardiovascular responses *Brain Res Bull*, 132:170-9.

29. Kim S J, Fong A Y, Pilowsky P M, Abbott SBG (2018) Sympathoexcitation following intermittent hypoxia in rat is mediated by circulating angiotensin II acting at the carotid body and subfornical organ *J Physiol*, 596:3217-32.

30. Kumar R, Thomas CM, Yong Q C, Chen W, Baker KM, et al. (2012) The intracrine renin-angiotensin system *Clin Sci (Lond)*, 123:273-84.

31. Lautner R Q, Villela D C, Fraga-Silva R A, Silva N, Verano-Braga T, Costa-Fraga F, Santos RA (2013) Discovery and characterization of alamandine: a novel component of the renin-angiotensin system *Circ Res*, 112:1104-11.

32. Leenen, FHH, Blaustein M P, Hamlyn JM (2017) Update on angiotensin II: new endocrine connections between the brain, adrenal glands and the cardiovascular system *Endocr Connect*, 6:R131-45.
33. Li H-B, Qin D-N, Ma L, Miao Y-W, Zhang D-M, Lu Y, Kang Y-M (2014) Chronic infusion of lisinopril into hypothalamic paraventricular nucleus modulates cytokines and attenuates oxidative stress in rostral ventrolateral medulla in hypertension *Toxicology and Applied Pharmacology*, 279:141-9.
34. Li P, Sun HJ, Cui BP, Zhou YB, Han Y (2013) Angiotensin-(1-7) in the rostral ventrolateral medulla modulates enhanced cardiac sympathetic afferent reflex and sympathetic activation in renovascular hypertensive rats *Hypertension*, 61:820-7.
35. Li W, Sullivan MN, Zhang S, Worker C J, Xiong Z, Speth R C, Feng Y, et al. (2015) Intracerebroventricular Infusion of the (Pro) renin Receptor Antagonist PRO20 Attenuates Deoxycorticosterone Acetate-Salt-Induced Hypertension *Hypertension* 65:352-61.
36. Lu P, Liang LW, Xu AL, Sun YY, Jiang S J, Shi Z (2020) Pro-inflammatory cytokines in the paraventricular nucleus mediate the adipose afferent reflex in rats *Pflugers Arch*, 472:343-54.
37. Ma H, Chen SR, Chen H, Pan HL (2019) Endogenous AT1 receptor-protein kinase C activity in the hypothalamus augments glutamatergic input and sympathetic outflow in hypertension *J Physiol*, 597:4325-40.
38. Marciante A B, Wang L A, Little J T, Cunningham J T (2020) Caspase lesions of PVN-projecting MnPO neurons block the sustained component of CIH-induced hypertension in adult male rats *Am J Physiol Heart Circ Physiol*, 318:34-48.
39. McKinley M J, Pennington G L, Ryan P J (2021) The median preoptic nucleus: A major regulator of fluid, temperature, sleep, and cardiovascular homeostasis *Handb Clin Neurol*, 179:435-54.
40. Mehay D, Silberman Y, Arnold AC (2021) The Arcuate Nucleus of the Hypothalamus and Metabolic Regulation: An Emerging Role for Renin-Angiotensin Pathways *Int J Mol Sci*, 22.
41. Miller A J, Arnold AC (2019) The renin-angiotensin system in cardiovascular autonomic control: recent developments and clinical implications *Clinical Autonomic Research*, 29:231-43.
42. Mohsin M, Souza LAC, Aliabadi S, Worker C J, Cooper SG, Afrin S, Feng Earley Y, et al. (2020) Increased (Pro)renin Receptor Expression in the Hypertensive Human Brain *Front Physiol*, 11-606811.
43. Mourão AA, de Mello ABS, Dos Santos Moreira M C, Rodrigues KL, Lopes PR, Xavier CH, Pedrino GR, et al. (2018) Median preoptic nucleus excitatory neurotransmitters in the maintenance of hypertensive state *Brain Res Bull*, 142:207- 15.
44. Mukerjee S, Gao H, Xu J, Sato R, Zsombok A, Lazartigues E, et al. (2019) ACE2 and ADAM17 Interaction Regulates the Activity of Presympathetic Neurons *Hypertension*, 74:1181-91.
45. Nasimi A, Haddad F, Mirzaei-Damabi N, Rostami B, Hatam M, et al. (2021) Another controller system for arterial pressure AngII-vasopressin neural network of the parvocellular paraventricular nucleus may regulate arterial pressure during hypotension *Brain Res* 1769:147618.
46. Natarajan, A, Jose, PA (2019) Renal Modulation: The Renin-Angiotensin-Aldosterone System (RAAS): Nephrology and Fluid/Electrolyte Physiology: Neonatology Questions and Controversies 107-27.
47. Pitra S, Worker CJ, Feng Y, Stern JE (2019) Exacerbated effects of prorenin on hypothalamic magnocellular neuronal activity and vasopressin plasma levels during salt-sensitive hypertension *Hypertension* 317:496-504.

48. Ren X, Zhang F, Zhao M, Zhao Z, Sun S, Fraidenburg D R, Han Y, et al. (2017) Angiotensin-(1-7) in Paraventricular Nucleus Contributes to the Enhanced Cardiac Sympathetic Afferent Reflex and Sympathetic Activity in Chronic Heart Failure Rats *Cell Physiol Biochem*, 42:2523-39.
49. Schelling P, Ganten U, Sponer G, Unger T, Ganten D (1980) Components of the Renin-Angiotensin System in the Cerebrospinal Fluid of Rats and Dogs with Special Consideration of the Origin and the Fate of Angiotensin II *Neuroendocrinology*, 31:297-308.
50. Schleifenbaum J (2019) Alamandine and Its Receptor MrgD Pair Up to Join the Protective Arm of the Renin-Angiotensin System *Front Med (Lausanne)*, 6:107.
51. Segiet A, Smykiewicz P, Kwiatkowski P, Żera T (2019) Tumour necrosis factor and interleukin 10 in blood pressure regulation in spontaneously hypertensive and normotensive rats *Cytokine*, 113:185-194.
52. Seki Y, Ichihara A, Mizuguchi Y, Sakoda M, Kurauchi-Mito A, Narita T, Itoh H, et al. (2010) Add-on blockade of (pro)renin receptor in imidapril-treated diabetic SHRsp 2:972-9.
53. Shen YH, Chen XR, Yang CX, Liu BX, Li P, et al. (2018) Alamandine injected into the paraventricular nucleus increases blood pressure and sympathetic activation in spontaneously hypertensive rats *Peptides* 103:98-102
54. Shi P, Diez-Freire C, Jun JY, Qi Y, Katovich MJ, Li Q, Raizada MK, et al. (2010) Brain microglial cytokines in neurogenic hypertension *Hypertension*, 56:297-303.
55. Shi Z, Jiang S-j, Wang G-h, Xu A-l, Guo L, et al. (2014) Pro-inflammatory cytokines in paraventricular nucleus mediate the cardiac sympathetic afferent reflex in hypertension *Autonomic Neuroscience*, 186:54-61.
56. Shi Z, Stornetta DS, Stornetta RL, Brooks VL (2022) Arcuate Angiotensin II Increases Arterial Pressure via Coordinated Increases in Sympathetic Nerve Activity and Vasopressin Secretion 9.
57. Simpson NJ, Ferguson AV (2017) The proinflammatory cytokine tumor necrosis factor- α excites subfornical organ neurons *Neuroscience* 118:1532-41.
58. Simpson NJ, Ferguson AV (2018) Tumor necrosis factor- α potentiates the effects of angiotensin II on subfornical organ neurons *Neuroscience* 315:R25-33.
59. Song CY, Khan NS, Liao FF, Wang B, Shin JS, Bonventre JV, Malik KU, et al. (2018) Brain Cytosolic Phospholipase A2 α Mediates Angiotensin II-Induced Hypertension and Reactive Oxygen Species Production in Male Mice *Am J Hypertens*, 31:622-9.
60. Souza LAC, Worker CJ, Li W, Trebak F, Watkins T, Gayban AJB, Feng Y, et al. (2019) (Pro)renin receptor knockdown in the paraventricular nucleus of the hypothalamus attenuates hypertension development and AT(1) receptor-mediated calcium events *Am J Physiol Heart Circ Physiol*, 316:1389-405.
61. Sriramula S, Cardinale JP, Francis, J (2013) Inhibition of TNF in the Brain Reverses Alterations in RAS Components and Attenuates Angiotensin II-Induced Hypertension *PLoS One*, 8:63847.
62. Stocker SD, Wenner MM, Farquhar WB, Browning KN (2022) Activation of the Organum Vasculosum of the Lamina Terminalis Produces a Sympathetically Mediated Hypertension *Hypertension*, 79:139-49.
63. Su Q, Huo CJ, Li HB, Liu KL, Li X, Yang Q, Kang YM, et al. (2017) Renin-angiotensin system acting on reactive oxygen species in paraventricular nucleus induces sympathetic activation via AT1R/PKC γ /Rac1 pathway in salt-induced hypertension *Sci Rep*, 7:43107.

64. Sun HJ, Li P, Chen WW, Xiong XQ, Han Y, et al. (2012) Angiotensin II and angiotensin- (1-7) in paraventricular nucleus modulate cardiac sympathetic afferent reflex in renovascular hypertensive rats *PLoS One*, 7:52557.
65. Tipnis SR, Hooper NM, Hyde R, Karran E, Christie G, Turner A J, et al. (2000) A human homolog of angiotensin-converting enzyme Cloning and functional expression as a captopril-insensitive carboxypeptidase *J Biol Chem*, 275:33238-43.
66. Trott DW, Harrison DG (2014) the immune system in hypertension 38:20-24.
67. Underwood CF, McMullan S, Goodchild AK, Phillips JK, Hildreth CM (2022) The subfornical organ drives hypertension in polycystic kidney disease via the hypothalamic paraventricular nucleus *Cardiovasc Res*, 118: 1138-49.
68. Wang F, Lu X, Liu M, Feng Y, Zhou S-F, Yang T, et al. (2015) Renal medullary (pro)renin receptor contributes to angiotensin II-induced hypertension in rats via activation of the local renin-angiotensin system *BMC Medicine*, 13:278.
69. Wang G, Woods C, Johnson MA, Milner TA, Glass MJ (2022) Angiotensin II Infusion Results in Both Hypertension and Increased AMPA GluA1 Signaling in Hypothalamic Paraventricular Nucleus of Male but not Female Mice *Neuroscience*, 485:129-44.
70. Wei SG, Yu Y, Felder RB (2018) Blood-borne interleukin-1 β acts on the subfornical organ to upregulate the sympathoexcitatory milieu of the hypothalamic paraventricular nucleus *American journal of physiology Regulatory, integrative and comparative physiology*, 314:447-58.
71. Wei SG, Yu Y, Felder RB (2021) TNF- α -induced sympathetic excitation requires EGFR and ERK1/2 signaling in cardiovascular regulatory regions of the forebrain *Am J Physiol Heart Circ Physiol*, 320: 772-86.
72. Wei SG, Yu Y, Zhang Z H, Felder RB (2015) Proinflammatory cytokines upregulate sympathoexcitatory mechanisms in the subfornical organ of the rat *Hypertension*, 65:1126-33.
73. WHO (2021) Cardiovascular diseases (CVDs) Retrieved from [https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-\(cvds\)#:~:text=Cardiovascular%20diseases%20\(CVDs\)%20are%20the,%2D%20and%20middle%2Dincome%20countries.](https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-(cvds)#:~:text=Cardiovascular%20diseases%20(CVDs)%20are%20the,%2D%20and%20middle%2Dincome%20countries.)
74. Woods C, Marques-Lopes J, Contoreggi N H, Milner T A, Pickel V M, Wang G, Glass M J, et al. (2021) Tumor Necrosis Factor α Receptor Type 1 Activation in the Hypothalamic Paraventricular Nucleus Contributes to Glutamate Signaling and Angiotensin II-Dependent Hypertension *J Neurosci*, 41:1349-62.
75. Worker CJ, Li W, Feng CY, Souza LAC, Gayban AJB, Cooper SG, Feng Earley Y, et al. (2020) The neuronal (pro)renin receptor and astrocyte inflammation in the central regulation of blood pressure and blood glucose in mice fed a high-fat diet *Am J Physiol Endocrinol Metab*, 318:765-78.
76. Wright JW, Harding J W (1997) Important role for angiotensin III and IV in the brain renin-angiotensin system *Brain Res Brain Res Rev*, 25:96-124.
77. Xu J, Molinas AJR, Mukerjee S, Morgan DA, Rahmouni K, Zsombok A, Lazartigues E, et al. (2019) Activation of ADAM17 (A Disintegrin and Metalloprotease 17) on Glutamatergic Neurons Selectively Promotes Sympathoexcitation *Hypertension*, 73:1266-74.
78. Xu J, Sriramula S, Xia H, Moreno-Walton L, Culicchia F, Domenig O, Lazartigues E, et al. (2017) Clinical Relevance and Role of Neuronal AT(1) Receptors in ADAM17-Mediated ACE2 Shedding in Neurogenic Hypertension *Circ Res*, 121:43-55.
79. Yu X-J, Miao Y-W, Li H-B, Su Q, Liu K-L, Fu L-Y, Kang Y-M (2019) Blockade of Endogenous Angiotensin-(1-7) in Hypothalamic Paraventricular Nucleus Attenuates High Salt-Induced Sympathoexcitation and Hypertension *Neuroscience Bulletin*, 35:47-56.

80. Yu XJ, Zhao YN, Hou YK, Li HB, Xia WJ, Gao HL, Kang YM, et al. (2019) Chronic Intracerebroventricular Infusion of Metformin Inhibits Salt-Sensitive Hypertension via Attenuation of Oxidative Stress and Neurohormonal Excitation in Rat Paraventricular Nucleus *Neurosci Bull*, 35:57-66.

81. Yu Y, Cao Y, Bell B, Chen X, Weiss RM, Felder RB, Wei SG, et al. (2019) Brain TACE (Tumor Necrosis Factor- α -Converting Enzyme) Contributes to Sympathetic Excitation in Heart Failure Rats Hypertension, 74:63-72.