

RESEARCH ARTICLE

Bio-ethanol production from Raspberry by yeast in repeated batch

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Citation: Reza Robati, Elhamalsadat Mirahmadinejad (2022) Bio-ethanol production from Raspberry by yeast in repeated batch. Stechnolock J Case Rep 3: 101

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ABSTRACT

Bio-ethanol, which is considered the cleanest liquid fuel, can be a reliable replacement to fossil fuels. Fermentation of sugar components of plant materials produces it. Berries are thought to be a good source of fermentation products, as they have a high sugar content and contain a variety of nutrients. This research looked at how well the yeast "Saccharomyces cerevisiae" produced ethanol from raspberry (Rubus ideaus) in repeated batches. The total sugar concentration of raspberry juice was 42.6 g/l, according to the findings. The maximum productivity, ethanol yield, and final bio-ethanol percentage were 6 g/l/h (g ethanol per liter of raspberry juice per hour), 35 g/l (g ethanol per liter of raspberry juice), and 7 g/l (g ethanol per liter of raspberry juice) respectively.

Keywords: Rubus ideaus Bio-fuels; Ethanol; Saccharomyces cerevisiae

Introduction

Due to the paucity of fossil fuels, more and more efforts have been made to produce alternate energy from renewable sources via microbial fermentation [17]. The conversion of biomass into fuel ethanol, the cleanest liquid fuel alternative to fossil fuels, has received a lot of attention [13]. Up to the 1930s, biologically produced alcohols, most notably ethanol, were utilized as a vehicle fuel [20]. Because of the abundant and inexpensive availability of fuel from petroleum and natural gas after WWII, however, there was little interest in employing agricultural crops for liquid fuel production [13]. Ethanol is made through the fermentation of sugars, starches, or cellulose by microorganisms and enzymes [11]. In Canada and USA, the raspberry (Rubus ideaus) is a popular fruit with numerous native cultivars [11]. Furthermore, the raspberry is Canada's most frequently farmed horticultural crop, with yearly production exceeding 4 million tons [9]. However, enormous volumes of raspberries are discarded as agricultural waste or sold as sub-standard quality on the market every year. [14]. As a result, a new product made from raspberry trash is particularly environmentally friendly. Because of its high sugar and nutritional content, the common raspberry is a good source of fermentation products. Raspberry bioethanol could be used as an advantageous substrate for acetic fermentation to generate raspberry vinegar. This vinegar has a greater mineral and organic acid content [14]. Although the manufacturing of gasoline ethanol is already an energy-efficient process, further research is being conducted to increase its long-term economic viability [9]. A yeast-based repeating batch procedure is widely regarded as a promising technology for effective alcohol synthesis, easy cell recycling, and stable operation [11]. Because useless raspberries are seasonal agricultural crops with limited supply, a repeated batch method rather than a continuous process is preferable in terms of operability for raspberry alcohol manufacturing [17].

Materials and Methods

Strain and Medium

Saccharomyces cerevisiae was taken from the Persian Type Culture Collection (PTCC) of the Iranian Investigate Organization for Science and Innovation. The strain was maintained on agar inclines containing YPD medium and the inoculums societies were vaccinated from the yeast on the slant by implies of sterile immunization circles. YPD medium containing 1 g glucose/dextrose, 1 g yeast extricate, and 2 g peptone per 100 ml refined water, was utilized for the inoculums medium but as expressed. Raspberry from Rasht city was utilized as the crude fabric for Alcohol fermentation. Raspberries were cut and squeezed in a mechanical juicer. The extract was collected in a glass vial (1,000 ml) and autoclaved at 120 °C for 20 min. The autoclaved juice was cooled and after that sifted utilizing Whatman (No. 1) channel paper.

Experiments

Raspberry juice for bio-ethanol generation was gotten as follows. Raspberry was squeezed in a mechanical juicer and the extract was instantly autoclaved for 30 min at 120 °C. The extract was at that point sifted twice, utilizing 6.0 pm pore-size membrane to expel coarse particles and a 0.4 pm pore-size membrane to expel organisms. By this method, 60 wt% of the raspberries prepared was recouped as raspberry juice. The filtered juice was put away in a fridge at 4.0 °C and utilized as needed. At that point S. cerevisiae was refined within the YPD medium broth within the shaker hatchery (VS-8480SF, Korea) at a temperature of 30 °C at 120 rpm for 24 h. The development of yeast was measured by spectrophotometer (600 nm) [17]. Then 50 ml of the raspberry extricate and 5 ml of inoculums were added to the four Erlenmeyer jars (two jars for typical S. cerevisiae and two jars for hereditarily adjusted strain of S. cerevisiae, obtained from Japan. The Group experiment was at that point carried out in carafes fitted with air circulation equipment. Alcohol aging was to begin with performed at 30 °C with CO_2 bubbling in arrange to preserve anaerobic condition. [15]. Four initial tests for yeast development were utilized at 30 °C for 48 h. Following this, tests for alcohol maturation were taken and kept at room temperature for 72 h. The ethanol product both by ordinary yeast and by a hereditarily adjusted strain was compared.

Analytical

Total organic acids in raspberry juice were measured by High- performance fluid chromatography (HPLC, 8020 model, Tosoh CO., Tokyo, Japan) and free amino acids were measured by the NBD-F (4-Fluoro-7-nitrobenzofurazan) strategy in combination with HPLC. A few minerals were decided by inductively coupled plasma nuclear emanation spectroscopy (ICP, Hitachi, Japan). The sum of sugars was measured by the phenol–sulphuric acid strategy [10]. Along these lines, fermentation tests were centrifuged (7,000 rpm) at 4 °C for 6 min and the supernatant were utilized for the assurance of ethanol by HPLC.

Results and Discussion

Ethanol, both renewable and ecologically inviting, is leading to an emotional increment in its generation capacity. Among numerous microorganisms that have been abused for ethanol generation, S. cerevisiae remains the prime one [3]. In this consider, the whole sugar concentration of raspberry juice was 42.6 g/l; in other thinks about, it was found to vary from 35.7 to 47.4 g/l [5]. This appeared that the examined raspberry may be a great source for bio-ethanol generation, as ethanol is delivered by the activity of microorganisms and enzymes through the aging of sugars. In brief, the composition of raspberry juice came to be; Total organic acids were 882.22 mg/l, Free amino acids were 954.57 mg/l, K 784.9 mg/l, PO₄ 130 mg/l, Mg 114.5 mg/l, Ca 130.2 mg/l, Na 1 mg/l, Mn 0.82 mg/l, Zn 0.64 mg/l, Fe 0.69 mg/l and the sucrose, fructose and glucose concentrations of raspberry juice were 10.42, 29.76 and 26.01 g/l. pH was 3.2. The maximum efficiency, ethanol abdicate and last ethanol percentage were 6 g/l/h, 35 g/l and 87 %, separately. Our results show high rates of bio-ethanol generation by raspberry juice as compared with others (6.5 g/l/h, 30.6 g/l and 91.9 %, individually) [19]. Total organic acids and free amino acids were 882.22 and 954.57 mg/l, respectively. The amounts of free amino acids and minerals are important because they are utilized by yeast as supplements and ethanol production is firmly coupled with the development of yeast cells [18]. Beneath anaerobic conditions, the pyruvate is further reduced to ethanol with the discharge of CO₂. The normal yeast tube has more CO₂ over the ethanol with an ethanol yield of 43 g/l but the ethanol surrender of the altered yeast was 46 g/l, affirming that the ethanol generation of the modified strain has more noteworthy immaculateness. In terms of sugar and supplement substance, it was basic to use discarded raspberry as a source of medium as they had been rejected basically on account of their shape or estimate [16]. Ethanol generation by ordinary S. cerevisiae and a hereditarily altered strain of S. cerevisiae was compared. The altered strain had more CO, above the ethanol. The application of modified yeast to improve ethanol generation from glucose may be a key in many biofuels' generation programs [2].

References

1. Akbas MY, Stark BC (2016) Recent trends in bioethanol production from food processing byproducts. J Ind Microbiol Biotechnol 43:1593-609

2. Alper H, Moxley J, Nevoigt E, Fink GR, Stephanopoulos G, et al. (2006) Engineering yeast transcription machinery for improved ethanol tolerance and production. Science 314:1565-8

3. Anderson S, Rahman PK (2018) Bioprocessing requirements for bioethanol: sugarcane vs. sugarcane bagasse. Handbook of Research on Microbial Tools for Environmental Waste Management 48-56

4. Barak S, Rahman RK, Neupane S, Ninnemann E, Arafin F, Laich AAC, Terracciano SS, et al. (2020) Measuring the effectiveness of high-performance Co-Optima biofuels on suppressing soot formation at high temperature. Proc. Natl. Acad. Sci. Unit. States Am 117: 3451-60

5. Bobinait eR, Viškelis P, Venskutonis PR (2016) Chemical composition of raspberry (Rubus spp.) cultivars In Nutritional Composition of Fruit Cultivars. Elsevier: Amsterdam, The Netherlands 713-31.

6. del Río PG, Gullón P, Rebelo F, Romaní A, Garrote G, Gullón B, et al. (2020) whole-slurry fermentation approach to high-solid loading for bioethanol production from corn stover Agronomy, 10: 1790

7. Ibeto A, Ofoefule K, Agbo (2011) A global overview of biomass potentials for bioethanol production: a renewable alternative fuel Trends 6: 410

8. Kalemba-Drożdż M, Kwiecień I, Szewczyk A, Cierniak A, Grzywacz-Kisielewska A, et al. (2020) Fermented Vinegars from Apple Peels, Raspberries, Rosehips, Lavender, Mint, and Rose Petals: The Composition, Antioxidant Power, and Genoprotective Abilities in Comparison to Acetic Macerates, Decoctions, and Tinctures. *Antioxidants* 9:1121

9. Luís Â, Domingues F, Pereira L (2018) Association between berries intake and cardiovascular diseases risk factors: A systematic review with meta-analysis and trial sequential analysis of randomized controlled trials Food Funct, 9:740-57

10. Masukot T, Minami A, Iwasaki N, Majima T, Nishimura SI, Lee YC, et al. (2005) Carbohydrate analysis by a phenol-sulfuric acid method in microplate format. Analytical Biochem 339:69-72

11. Morales CG, Pino MT, del Pozo A (2013) Phenological and physiological responses to drought stress and subsequent rehydration cycles in two raspberry cultivars. Sci. Hortic 162: 234-41.

12. Naghshbandi MP, Tabatabaei M, Aghbashlo M, Gupta VK, Sulaiman A, Karimi K, Moghimi H, Maleki M, et al. (2019) Progress toward improving ethanol production through decreased glycerol generation in Saccharomyces cerevisiae by metabolic and genetic engineering approaches Renew. Sustain. Energy Rev., 115: 109353

13. Nouri H, Ahi M, Azin M, Gargari SLM (2020) Detoxification vs. adaptation to inhibitory substances in the production of bioethanol from sugarcane bagasse hydrolysate: a case study Biomass Bioenergy, 139: 105629

14. Pan P, Kang S, Wang Y, Liu K, Oshima K, Huang YW, Zhang J, Yearsley M, Yu J, Wang LS, et al. (2017) Black Raspberries Enhance Natural Killer Cell Infiltration into the Colon and Suppress the Progression of Colorectal Cancer. Front. Immunol, 8:997

15. Robati R. (2013) Bio-ethanol production from green onion by yeast in repeated bath. Indian J Microbiology. 53:329-31

16. Robati R, Fathi A and gholami S (2013) Bio-ethanol production from Ajowan plant. Annals of biological research, 4: 7-11.

17. Sanda T, Hasunuma T, Matsuda F, Kondo A (2011) Repeated-batch fermentation of lignocellulosic hydrolysate to ethanol using a hybrid Saccharomyces cerevisiae strain metabolically engineered for tolerance to acetic and formic acids. Bioresour Technol 102:7917-24

18. Siti Hajar M, Rahmath A, Siti Azmah J, Hartinie M, Jualang G, Ainol Azifa Mohd Faik, Kenneth R, et al. (2017) Yeasts in sustainable bioethanol production: A review, Biochemistry and Biophysics Reports, 10:55

19. Vazirzadeh M and Robati R (2013) Investigation of bio-ethanol production from waste potatoes. Annals of biological research, 4:104-06.

20. Vohra M, Manwar J, Manmode R, Padgilwar S, Patil S, et al. (2014) Bioethanol production: feedstock and current technologies. Journal of Environmental Chemical Engineering, 2:573-84